



Plasmid DNA concentration and desalination

User manual

NucleoSpin[®] Finisher Midi

February 2016 / Rev. 01

Plasmid DNA concentration and desalination

Protocol-at-a-glance (Rev. 01)

NucleoSpin® Finisher Midi


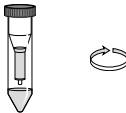

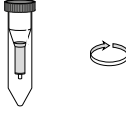
1 Adjust precipitation conditions		2.5 mL Buffer FB to 5 mL anion exchange eluate Vortex for 5 s
2 Filtrate		Load mixture 1 min, 3,000 x <i>g</i> Discard flow-through
3 Wash and dry filter membrane		2 mL Buffer A4 2 min, \geq 3,000 x <i>g</i>
4 Elute DNA		200–500 μ L H ₂ O-EF RT, 1 min 2 min, \geq 3,000 x <i>g</i>

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1 Components

1.1 Kit contents

REF	NucleoSpin® Finisher Midi	
	10 preps 740439.10	50 preps 740439.50
Buffer FB	30 mL	150 mL
Buffer A4 (Concentrate)*	6 mL	25 mL
H ₂ O-EF	13 mL	30 mL
NucleoSpin® Finisher Midi Columns (in 50 mL Collection Tubes)	10	50
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1.2 Reagents and equipment to be supplied by user

Reagents

- 96–100 % ethanol

Equipment

- Centrifuge capable of reaching $\geq 3,000 \times g$ with adaptors for 50 mL centrifuge tubes
- Pipettes and pipette tips for 0.1–1 mL and 0.5–10 mL

* For preparation of working solution and storage conditions see section 3.

2 Product description

2.1 Basic principle

NucleoSpin® Finisher kits supersede the commonly used and time-consuming isopropanol precipitation of DNA from anion exchange eluates by allowing the rapid DNA precipitation onto a filter membrane, followed by a combined washing / drying step and a convenient elution with a variable elution volume. All steps are conveniently performed in a centrifuge which allows the parallel purification of multiple samples.

DNA is precipitated by Buffer FB and filtered by the specially designed matrix of the novel **NucleoSpin® Finisher Columns**, followed by a combined washing and drying step with ethanolic Wash Buffer A4 to remove salts, impurities, and ethanol.

Afterwards, plasmid DNA can be eluted in supplied endotoxin-free H₂O-EF. The applied elution volume can be adjusted from 100 µL to 1.5 mL according to the expected amount of plasmid DNA to ensure optimal yield and concentration (see section 2.4 for recommendations concerning the optimal elution volume).

2.2 Kit specifications

The **NucleoSpin® Finisher Midi** kit is specifically designed for the fast and parallel purification and concentration of up to 2 mg of plasmid DNA from anion exchange preparation eluates. The **NucleoSpin® Finisher Midi** kit can be used in combination with the **NucleoBond® Xtra Midi** and the **NucleoBond® PC 100** kits.

Purification of DNA from maxi prep eluates (**NucleoBond® Xtra Maxi** and **NucleoBond® PC 500**) is also possible, but will require multiple loading steps and additional precipitation buffer (to be ordered separately).

2.3 Recommendations for pipetting of Buffer FB

! Buffer FB is viscous. Use of **reverse pipetting** is recommended to ensure accurate volumes. Reverse pipetting is done by pressing down the pipette's plunger button all the way down to the second stop before slowly aspirating the Buffer FB until the plunger button rests again in the starting position. The buffer volume inside the pipette tip is larger than set now, so when dispensing the Buffer FB to the anion exchange eluates, be sure to dispense to the first stop only! Liquid remaining in the pipette tip can be dispensed back to the original solution.

For further details concerning the reverse pipetting technique and liquid handling of viscous fluids, you may also check your pipette manufacturer's information material.

2.4 Elution procedures

Total yield depends on the final DNA concentration in the eluates.

A higher concentration than 2 µg/µL of DNA is highly viscous and therefore difficult to elute from a spin column. As a result, DNA will not elute completely if the elution buffer volume is too low.

To prevent reduced total yield as side-effect of high concentration, it is recommended to measure the DNA content of the anion exchange eluate and to choose the total elution volume accordingly to gain a final concentration of 1–2 µg/µL.

- For a high **concentration**, use an elution volume of 200 µL, reload the eluate onto the column, and repeat the elution step.
- For a high **yield**, use an elution volume of 500 µL, reload the eluate onto the column, and repeat the elution step.

In general, it is advantageous to incubate the Elution Buffer on the membrane at room temperature or elevated temperatures (e.g., 50–70 °C) for 1–5 min.

Recommended elution volumes according to expected yield		
Kit	DNA yield	Recommended elution volume
NucleoBond® PC 100	up to 100 µg	100–150 µL
NucleoBond® Xtra Midi	up to 500 µg	200–500 µL
NucleoBond® PC 500	up to 500 µg	200–500 µL
NucleoBond® Xtra Maxi	up to 1500 µg	2 x 500 µL

3 Storage conditions and preparation of working solutions

All kit components can be stored at room temperature (18–25 °C) and are stable for at least two years.

Before starting any **NucleoSpin® Finisher Midi** protocol, prepare the following:

- **Wash Buffer A4:** Add the given volume of ethanol (96–100 %) as indicated on the bottle or in the table below to **Buffer A4 Concentrate** before first use. Mark the label on the bottle to indicate that the ethanol is added. Prepared Buffer A4 is stable at room temperature (18–25 °C) for at least one year.

NucleoSpin® Finisher Midi		
REF	10 preps 740439.10	50 preps 740439.50
Buffer A4 (Concentrate)	6 mL Add 24 mL ethanol	25 mL Add 100 mL ethanol

4 Safety instructions

The NucleoSpin® Finisher Midi kit does not contain hazardous components.

5 Protocol for plasmid concentration and desalination

5.1 Plasmid purification from anion exchange prep eluates

Before starting the preparation:

Check if Buffer A4 was prepared according to section 3.

1 Adjust precipitation conditions

Add **2.5 mL Buffer FB** to **5 mL** anion exchange eluate.

Reverse pipetting is recommended (see section 2.3).

Vortex for **5 s**.

2 Filtrate

Load the mixture onto a **NucleoSpin® Finisher Midi Column** combined with a 50 mL Collection Tube.

Centrifuge for **1 min** at **3,000 x g**. If the filtration is not complete, repeat centrifugation until all fluid has passed the filter membrane.

Discard flow-through and place the NucleoSpin® Finisher Midi Column back into the empty Collection Tube.

3 Wash and dry filter membrane

Apply **2 mL Buffer A4** onto the column. Centrifuge for **2 min** at **≥ 3,000 x g**. Discard collection tube and place the NucleoSpin® Finisher Midi Column into a new 50 mL Collection Tube (not supplied).

4 Elute DNA

Add **200–500 µL** (minimum 100 µL, maximum 1.5 mL) of **nuclease- and endotoxin-free H₂O-EF** onto the membrane and incubate at **room temperature** for **1 min**.

Centrifuge for **2 min** at **≥ 3,000 x g**.

Optional: Repeat elution with the eluate as elution buffer for optimal recovery. See section 2.4 for further recommendations.

5.2 DNA precipitation from maxi prep eluates

Note: The supplied volume of Buffer FB in this kit will not be sufficient for purification of maxi prep eluates. Additional buffer has to be ordered separately (see ordering information).

Add **7.5 mL Buffer FB** to **15 mL** anion exchange maxi prep eluate. Load up to 8 mL of the mixture onto a **NucleoSpin® Finisher Midi Column** combined with a 50 mL Collection Tube. Centrifuge for **1 min** at **3,000 x g**. If the filtration is not complete, repeat centrifugation until all fluid has passed the filter membrane.

Discard flow-through and place the NucleoSpin® Finisher Midi Column back into the empty collection tube. Repeat this step until all the mixture from step 1 has passed the filter membrane.

Continue with step 3 of the standard protocol.

6 Appendix

6.1 Troubleshooting

Problem	Possible cause and suggestions
	<p><i>No plasmid DNA present in anion exchange eluates.</i></p> <ul style="list-style-type: none"> • Measure DNA yield after anion exchange prep.
No or low DNA yield	<p><i>Insufficient amount of Buffer FB added.</i></p> <ul style="list-style-type: none"> • Buffer FB is viscous, make sure to add the correct volume. • Use “reverse pipetting” to avoid inaccurate pipetting of precipitation buffer (see section 2.3). • Precipitation works best when 0.5 vol of Buffer FB are added to each vol of anion exchange eluate. When using other volumes than those of the standard procedure, adjust volume of Buffer FB accordingly.

6.2 Ordering information

Product	REF	Pack of
NucleoSpin® Finisher Midi	740439.10	10 preps
	740439.50	50 preps
MN Vacuum Manifold	740299	1
NucleoSnap Finisher Midi	740434.10	10 preps
	740434.50	50 preps
NucleoSnap Finisher Maxi	740435.10	10 preps
	740435.50	50 preps
Buffer FB	740438.1000	1000 mL
Buffer A4 (Concentrate)	740914.1	200 mL
H ₂ O-EF	740798.1	1000 mL

6.3 Product use restriction/warranty

NucleoSpin® Finisher Midi kit components are intended, developed, designed, and sold FOR RESEARCH PURPOSES ONLY, except, however, any other function of the product being expressly described in original MACHEREY-NAGEL product leaflets.

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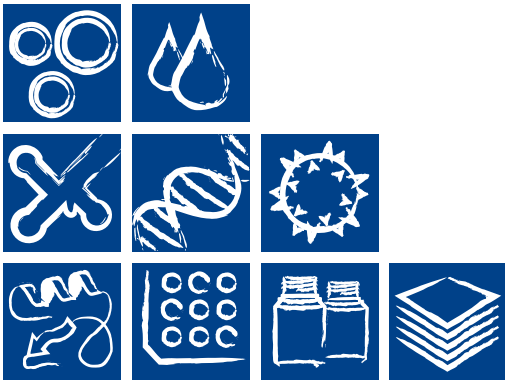
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